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Transcriptome analysis of *Streptococcus gordonii* Challis DL1
indicates a role for the biofilm-associated *fruRBA* operon in
response to *Candida albicans*

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SUMMARY

Multiple levels of interkingdom signaling have been implicated in maintaining the ecological balance between *Candida albicans* and commensal streptococci to assure a state of oral health. To better understand the molecular mechanisms involved in the initial streptococcal response to the presence of *C. albicans* that can initiate oral surface colonization and biofilm formation, hypha-forming cells were incubated with *Streptococcus gordonii* cells for 30 minutes to assess the streptococcal transcriptome response. A genome wide microarray analysis and quantitative PCR validation of *S. gordonii* transcripts identified a number of genes, the majority of which were involved in metabolic functions that were differentially expressed in the presence of hyphae. The *fruR*, *fruB* and *fruA* genes encoding the transcriptional regulator, fructose-1-phosphate kinase, and fructose-specific permease, respectively, of the phosphoenolpyruvate-dependent fructose phosphotransferase system, were consistently up-regulated. An *S. gordonii* mutant in which these genes were deleted by allelic replacement, formed an architecturally-distinct, less robust biofilm with *C. albicans* than did parental strain cells. Complementing the mutant with plasmid borne *fruR*, *fruB* and *fruA* genes caused phenotype reversion, indicating that the genes in this operon played a role in dual species biofilm formation. This genome wide analysis of the *S. gordonii* transcriptional response to *C. albicans* has identified several genes that have potential roles in interkingdom signaling and responses.

INTRODUCTION

Establishment of multispecies oral biofilms involves specific physicochemical adhesin-receptor interactions that can facilitate cell attachment and coaggregation as well as intercellular communication between microorganisms (Jenkinson, 2011; Wright *et al.*, 2013). These interactions can be mediated by diffusible signals and metabolic interactions that affect the microenvironment within a developing biofilm (Kolenbrander *et al.*, 2010; Nobbs & Jenkinson, 2015). Recent studies have provided insights into the many levels of cross talk between commensal oral streptococcal species and *Candida albicans* cells that cohabit the same niche in the oral cavity (Morales & Hogan, 2010; Xu *et al.*, 2014a; Bertolini *et al.*, 2015; Jack *et al.*, 2015). Balances in growth between these two genera (Diaz *et al.*, 2012) are thought to be essential in preventing *Candida* opportunistic infections and maintaining oral health. Contrarily, there are synergistic interactions between streptococci and *C. albicans* that have been shown to enhance fungal (Xu *et al.*, 2014b) or bacterial (Falsetta *et al.*, 2014) pathogenicity.

Studies of *Candida*-streptococcal interactions suggest that oral *Streptococcus gordonii*, a model commensal oral streptococcal species, releases diffusible signals and metabolic by-products that influence *C. albicans* hypha formation and dual species biofilm development (Bamford *et al.*, 2009). On the other hand, proteomic (Peters *et al.*, 2010) and transcriptomic studies (Sztajer *et al.*, 2014) have shown that the presence of *C. albicans* can result in changes in cumulative protein and transcript levels in Gram-positive bacterial species after several hours of co-cultivation.

To better understand the molecular mechanisms involved in the initiation of *C. albicans*-streptococcal biofilm formation, the present studies focused on the streptococcal short-term response to the presence of *C. albicans* under hyphae-forming conditions. Microarray analysis

of the *S. gordonii* transcriptome identified a number of genes that were differentially expressed in response to hyphae. There was consistent up-regulation of the *fruR*, *fruB* and *fruA* genes which constitute the operon expressing a fructose-specific phosphotransferase. This operon has previously been shown to influence *S. gordonii* biofilm formation (Loo *et al.*, 2003). Accordingly, the role of these genes in early dual species biofilm formation was explored utilizing a *S. gordonii* $\Delta fruRBA$ mutant strain in which the operon was deleted by allelic replacement. The results indicate that the *fruRBA* operon, and a range of other metabolic genes, are differentially expressed in *S. gordonii* in response to the presence of *C. albicans* hyphae. Furthermore, the *fruR*, *fruB* and *fruA* genes are implicated in the initiation and architecture of *C. albicans*-streptococcal biofilms.

METHODS

Interaction of strain DL1 with *C. albicans* hyphae

S. gordonii Challis DL1 cells were grown to mid-log phase (approximately 10^8 cells ml⁻¹) in TY medium (5 g l⁻¹ Tryptone, 5 g l⁻¹ yeast extract, 23 mM K₂HPO₄, pH 7.5) supplemented with 5 g l⁻¹ glucose (TY-G) under stationary conditions at 37°C under 5% CO₂. *Candida albicans* SC5314 cells were grown in YPD medium (1% yeast extract, 2% mycological peptone, 2% glucose) aerobically with shaking at 225 rpm, for 16 h at 30°C (OD₆₀₀ ~2.6). The *C. albicans* cells were harvested by centrifugation (1,370 x *g* for 5 min), washed once in complete salts-biotin (CSB) nutrient-limited medium (7.5 mM ammonium sulfate, 15 mM KH₂PO₄, 0.1 mM MgSO₄·7H₂O, 3.6 µg ml⁻¹ biotin, 0.05 mg ml⁻¹ CaCl₂, pH 6.5), and suspended in fresh CSB at OD₆₀₀ = 1.0 (~1 x 10⁷ cells ml⁻¹). The culture was incubated aerobically with 225 rpm-shaking for 16 h at 37°C to induce starvation (Holmes & Shepherd, 1988). *C. albicans* hyphae formation was then triggered by the addition of one-tenth volume 10% glucose (CSB-Glc) to the *C. albicans* culture, and

incubation with shaking was continued for 3 h at 37°C, at which time the pH was measured. Under these conditions at least 80% of the cells were forming hyphal filaments (germ tubes) (Jenkinson *et al.*, 1990).

Equal volumes of the *S. gordonii* and *C. albicans* cultures, or *S. gordonii* with final pH-adjusted (see above) fresh CSB-Glc medium as control, were mixed, centrifuged (3,800 x *g* for 5 min) and the cell pellets were incubated without shaking for 30 min at 37° C aerobically (in 5% CO₂) to allow the *S. gordonii* cells to interact with the *C. albicans* hyphae in close proximity. The *S. gordonii*-CSB-Glc control adjusted to the pH of the *C. albicans* hyphae-forming culture took into account any effects of centrifugation, and the shift in both nutritional status and pH for *S. gordonii* when mixed with *C. albicans*. At the end of this co-incubation, total RNA was extracted from the cell pellets. For microarray transcriptome analyses a minimum of two biologically independent *S. gordonii* + *C. albicans* hyphae interaction experiments were performed.

To verify the specificity of *S. gordonii* gene responses to growing hyphae, two equivalent volumes of hyphae were harvested by centrifugation (3800 x *g*) and one aliquot was suspended in PBS (10 mM Na₂HPO₄, 1.8 mM KH₂PO₄, pH 7.4 containing 0.137 M NaCl and 2.7 mM KCl) and heated for 60 min at 100°C to induce killing. Killed or live hyphae suspensions, as well as a suspension of an equivalent number of cells of the irrelevant bacterium *Enterococcus faecalis* strain OG1x as a control, were each resuspended with an equivalent number of *S. gordonii* cells in pH-adjusted CSB, re-centrifuged and co-incubated as above. Two independent biological replicate experiments were performed.

RNA isolation and cDNA synthesis

RNA was prepared from the *S. gordonii* + *C. albicans* hyphae cell mixtures or *S. gordonii* + CSB-Glc control with a FastPrep120 Instrument (MP Biomedicals, Santa Ana, CA, USA) and RNAPro Blue RNA Extraction Kit (MP Biomedicals) for prokaryotic cells according to the

manufacturer's directions. The extracted RNA was digested with DNase (Roche Applied Science, Indianapolis, IN, USA) and purified with QIAgen RNA mini spin columns (QIAgen, Germantown, MD, USA) according to the manufacturer's directions. RNA integrity was confirmed by comparison of the ratios of absorbance at 260/280 nm and 260/230 nm as quantified using a NanoDrop spectrophotometer (Thermo Fisher Scientific, Pittsburgh, PA, USA). For real-time PCR experiments cDNA was reverse transcribed from the purified RNA using the iScript cDNA Synthesis Kit (BioRad, Hercules, CA, USA) according to the manufacturer's directions. For microarray experiments cDNA was reverse transcribed using Superscript III (Life Technologies, Grand Island, NY, USA), end-labeled via amino-allyl-labeling (Life Technologies, Grand Island, NY, USA) and Cy5-dye coupled (GE Healthcare/Amersham, Piscataway, NY, USA) to make microarray probes as previously described (Vickerman *et al.*, 2007). Probes were purified over QIAgen PCR purification columns (QIAgen) to remove excess dye before use.

Microarray hybridization

For microarray experiments, cDNA probes were hybridized to aminosilane-coated glass slides spotted with unique 70-mers representing each open reading frame from the *S. gordonii* strain CH1 genome sequence (Vickerman *et al.*, 2007; Genbank number CP000725.1) in replicates of six (obtained from the Pathogen Functional Genomics Resource Center; NIDCR Oral Microbe Microarray Initiative). Each slide contained a total of 500 *Arabidopsis thaliana* 70-mers as controls. An *in silico* analysis of the *A. thaliana* and 2,076 *S. gordonii* probes via a BLAST-N algorithm (Altschul *et al.*, 1990) returned no significant similar sequences in the *C. albicans* SC5314 genome (AACQ000000000). Control experiments using cDNA prepared from pure *C. albicans* hyphal cells showed no specific hybridization of *C. albicans* cDNA to any of the *S. gordonii* spotted oligonucleotides.

Slides were blocked during pre-hybridization at 42°C in 5x SSC blocking buffer (1x SSC is 0.015 M sodium citrate, pH 7.4, containing 0.15 M NaCl, 0.1% SDS, and 1% bovine serum albumin), then washed successively with water and isopropanol before drying via centrifugation prior to hybridization. To accommodate potential increases in cDNA quantity based upon contributions from the *C. albicans* hyphae, two-fold more cDNA from the dual species interaction samples was mixed with *S. gordonii* + CSB-Glc control samples for each interaction comparison. To account for biases during the labeling process, flip-dyes, in which the fluorescent labeling was reversed for each sample, were used (Quackenbush, 2002). Cy3-labeled *S. gordonii* CSB-Glc cDNA was combined with Cy5-labeled dual species cDNA in appropriate ratios; similarly, Cy5-labeled *S. gordonii*-CSB-Glc and Cy3-labeled dual species cDNAs were combined. The dye-labeled cDNA mixtures were dried, suspended in 50 µl hybridization buffer (40% v/v formamide, 5x SSC, 0.1% SDS, 0.6 mg ml⁻¹ Salmon sperm DNA), heated for 10 min at 95°C, placed on ice, and individual mixtures added to each microarray slide under a Lifterslip (Erie Scientific Company, Portsmouth, NH, USA). Each slide was incubated for 16 h at 42°C in a sealed hybridization chamber (Corning Life Sciences, Tewksbury, MA, USA), then washed successively in 0.1% SDS-2x SSC, 0.1% SDS-0.1x SSC, and 0.1x SSC, and dried by centrifugation.

Microarray analysis

The Axon 4200A (Axon Instruments, Foster City, CA, USA) was used to obtain digital images of the microarrays. Data were analyzed using the TM4 Microarray Software Suite, consisting of Spottfinder, Midas, and MeV (<http://www.tm4.org/>) (Saeed *et al.*, 2003). The fluorescence signals from the flip-dye hybridizations were measured using Spottfinder. Normalization and regularization of the data were performed using Midas (Lowess normalization, block mode, smooth parameter of 0.33, Cy3 reference; standard deviation regularization, block mode, Cy3 reference; flip-dye consistency checking and combination, using the data trim option of the

standard deviation mode and keeping all data within 2.0 standard deviations; replicate spots were averaged to be used in the determination of the final ratio for each spot). Using MeV, with 100 permutations and a *P* value cutoff of 0.05 the final expression ratios were generated and used to determine the fold change for hybridization of each probe; fold changes were ranked from the greatest increase to the greatest decrease. Experimental cDNA probes from *S. gordonii* cells incubated with hyphae that hybridized to the oligonucleotide spots with average fold changes 1.6-times greater than or 1.6-times less than fold changes of *S. gordonii*-CSB-Glc controls in two biologically independent experiments were considered to be potentially up- or down-regulated, respectively.

Data deposit

Microarray data have been deposited in NCBI's Gene Expression Omnibus (<http://www.ncbi.nlm.nih.gov/geo/>) and are available through GEO Series accession number: GSE68752. The files are linked to Genbank entry CP000725.1. Currently, a new annotation of the *S. gordonii* Challis genome is in progress (NC_009785) using the annotation method of Tatusova *et al.* (2014). A comparison of original and updated locus tags is included in supplemental Table S2.

Real-time PCR and data analysis

Qualitative real-time PCR (qPCR) assays were performed to confirm the expression of selected *S. gordonii* DL1 genes that appeared to be up- or down-regulated in response to the presence of hyphae, based upon the microarray results. Primers for specific genes were designed using Primer3, available online at <http://primer3.ut.ee/> (Koressaar & Remm, 2007; Untergrasser *et al.* 2012) to generate 100- or 150-bp amplicons as required (Table S1). Specificity of the primers was confirmed by searching for nucleotide sequence similarity in the *S. gordonii* genome (CP00725.1). The qPCR was performed using the Applied Biosystems 7500 Real-Time PCR

System (Life Technologies) and carried out in 96-well microtiter plates using Power®SYBR® Green Master Mix (Life Technologies) according to the manufacturer's directions. A minimum of 3 replicates were assayed for each sample. Data were analyzed using SDS Software v1.4 (Life Technologies). Expression values were determined using the $2^{-\Delta\Delta C_t}$ method (Livak & Schmittgen, 2001).

Construction of *fruRBA* deletion and complemented strains

A derivative of the parental strain Challis DL1 with a deletion of the *fruRBA* chromosomal region was generated by a PCR ligation mutagenesis approach (Figure S1). Successive overlapping PCR products were made with primers listed in Table S1 using strain DL1 chromosomal template to amplify the DNA regions that flank the 5-prime and 3-prime ends of the *fru* operon with an intervening 1084-bp *aad9* gene encoding spectinomycin resistance. The resulting amplicon was a dsDNA fragment in which 3,659 bp of the *fruRBA* operon, including the putative -10 promoter region and predicted catabolite repressible element binding site (Loo *et al.*, 2003), were replaced with 1,084-bp DNA region encoding *aad9* with its native promoter and terminator from plasmid pFW5 (Podbielski *et al.*, 1996). This DNA fragment was transformed directly into serum-competent strain DL1 cells and transformant colonies, which arose via allelic replacement (Fig. S1), were selected by growth on agar medium supplemented with 100 µg ml⁻¹ spectinomycin. After confirmation of the expected chromosomal sequence by nucleotide sequence analysis, the resulting strain with a deletion of the *fruRBA* region was designated strain UB2683.

To complement the mutation, a 3,984-bp amplicon was made from strain DL1 chromosomal template with primers BamFruCompF and BamFruCompR (Table S1) by PCR using Elongase enzyme (Lifetech). The resulting DNA encoding the complete *fruRBA* operon (Fig. S1) was digested with *Bam*H1 and ligated into an engineered *Bam*H1 site (Vickerman *et al.* 2003) of the streptococcal replicative plasmid pVA749 (Macrina *et al.*, 1981) and transformed

into the parental strain. The sequence fidelity of the cloned fragment was confirmed by nucleotide sequence analysis and the resulting streptococcal plasmid designated p49MfruRBA was transformed into strain UB2683. Putative transformants carrying the plasmid were selected on Todd Hewitt agar medium with erythromycin ($5 \mu\text{g ml}^{-1}$) and a representative UB2683/p49MfruRBA transformant was chosen for further characterization and biofilm studies.

Preparation of saliva

Pooled human saliva, collected from a minimum of six volunteers as approved by the National Research Ethics Committee South Central Oxford C (Number 08/H0606/87+5), was treated with 0.25 M dithiothreitol, and clarified by centrifugation, as previously described (Dutton *et al.*, 2014). The resulting supernatant was diluted to 10% with sterile water, filter sterilized with 0.45 micron pore size membranes, stored at -20°C , and used to coat glass cover slips as substrata in biofilm growth experiments.

Single species biofilms

For mono-strain studies to determine the abilities of the *S. gordonii* parent or mutant cells to form biofilms on saliva-coated substrata, cells were grown in one of four different biofilm media: TY-G, TY medium with 5 g l^{-1} fructose (TY-F), and Yeast Nitrogen Base, Tryptone, KH_2PO_4 - K_2HPO_4 pH 7.0 (YPT) with either 1 g l^{-1} glucose (YPT-G) or 1 g l^{-1} fructose (YPT-F). Biofilms were grown on 19-mm glass cover slips contained within 12-well tissue culture plates. Sterile cover slips were incubated with 10% sterile saliva for 16 h at 4°C . Saliva was removed and 0.9 ml of either TY-G, TY-F, YPT-G or YPT-F medium was added. Streptococcal strains were grown for 16 h at 37°C in Brain Heart Infusion-Yeast Extract (BHY) medium, inoculated (1:10 dilution) into TY-G, TY-F, YPT-G or YPT-F medium, and incubated until $\text{OD}_{600} = 0.6$ was obtained. Culture aliquots (0.1 ml) were added to wells containing 0.9 ml of the corresponding fresh media and incubated in a candle jar for 16 h at 37°C . Cover slips were removed into a

fresh 12 well plate, washed twice with PBS, stained with 0.5% crystal violet, washed 3 times with PBS, air dried, and visualized by transmitted light microscopy (Leica DMLB). Alternatively, for biomass studies, following staining with crystal violet, cover slips were transferred to a fresh 12 well plate, and washed three times with PBS. The stain was then released by the addition of 10% acetic acid; the absorbance of the eluate was measured at 595 nm (Jakubovics *et al.*, 2005).

Dual species biofilms

Biofilms were grown in 35-mm diameter plastic culture dishes with 14-mm No. 1.0 base glass cover slip windows (Mat Tek). Cover slip windows were coated with 2 ml sterile saliva for 16 h at 4°C. Saliva was removed by aspiration and 1.8 ml YPT-G and 0.2 ml *C. albicans* cell suspensions (2×10^6 in YPT) was added to each dish. Dishes were incubated for 1 h at 37°C, with gentle shaking at 50 rpm in a humid environment, and then the unattached *C. albicans* cells were removed. For control biofilms (*C. albicans* only), fresh YPT-G medium was added. For dual species biofilms, and streptococcal biofilm controls, overnight cultures of *S. gordonii* cells were suspended at $OD_{600} = 0.5$ in YPT-G and 2 ml aliquots were added to the *C. albicans*-inoculated plates to initiate dual species biofilm formation, or to saliva only-coated cover slip windows (*S. gordonii* biofilm controls). After incubation for 30 min at 37°C, to allow streptococcal cell attachment, unattached cells were removed by aspiration, fresh YPT-G medium was added (2 ml) and incubation was continued aerobically for up to 5 h at 37°C. At the end of the experiment, the biofilm-coated cover slip windows were removed, rinsed with YPT, and either stained with crystal violet for examination by light microscopy, or stained with fluorescein isothiocyanate (FITC), rinsed, and examined by confocal laser scanning microscopy (CLSM) using a Leica SP5-AOBS confocal microscope attached to a Leica DM I6000 inverted epifluorescence microscope as previously described (Dutton *et al.*, 2014). Velocity® software was used to generate 3D images.

RESULTS

DL1 transcriptional changes in response to co-incubation with hyphae

S. gordonii genes found to be differentially expressed in *S. gordonii* DL1 cells co-incubated for 30 minutes with hyphae as compared to genes in strain DL1 cells incubated with control CSB-Glc medium are shown in Table 1. All genes listed exhibited an average fold change difference of at least 1.6 in two independent biological replicate experiments and met the significance cutoff of 95% confidence. Complete transcriptome data from these experiments have been deposited in the GEO database.

Microarray analysis indicated a consistent 3- to 4- fold upregulation of the *fruRBA* operon (Table 1) in strain DL1 in response to a 30-minute incubation with *C. albicans* hyphae. Results from each microarray experiment were validated by independently-synthesized cDNA by quantitative PCR (Table 2). All three genes in the *fruRBA* operon were up-regulated in strain DL1 cells that interacted with hyphae compared to control DL1 cells incubated with pH-adjusted CSB-Glc. Furthermore, the up-regulated response of these genes was specific to live hyphae (Table 3) since equivalent numbers of heat-killed hyphae or the unrelated bacterium *E. faecalis* did not induce *fruRBA* under the co-incubation conditions.

The microarray data also showed a consistent upregulation of locus SGO_0702 which encodes a small hypothetical protein of unknown function (Table 1). This locus overlapped the convergent locus SGO_0701 which encodes a putative DNA-binding histone-like HU protein. Due to the small size of these open reading frames (276 and 237 bp for SGO_0701 and SGO_0702, respectively) the microarray probes for the two loci overlapped each other by 64 of 70 nucleotides. Nevertheless, the unidirectionality of the cDNA probes used in the microarray analysis allowed specific detection of transcripts arising from SGO_0702, which were up-regulated, versus the transcripts from the convergent SGO_0701 which was not differentially expressed. Two independent qPCR reactions were designed to validate these results (Table 2).

The primers for SGO_0701 were completely within the open reading frame and indicated no significant upregulation of this gene in response to hyphae. In contrast, qPCR results for SGO_0702 showed an increase in relative expression of this gene in strain DL1 cells that interacted with hyphae. However, due to the small size of the two loci and the limits of primer design, primer 702-left was within SGO_0702 but primer 702-right overlapped the 3-prime end of SGO_0701. Since qPCR requires bidirectional amplification of the transcript, it cannot be ruled out that some of the upregulation of SGO_0702 detected by qPCR is due to a contribution of read through of the transcript from SGO_0701. Overall, the increased expression of SGO_702 taken together with the microarray results suggests that there is increased transcription of this locus in *S. gordonii* DL1 cells that interacted with hyphae under the conditions tested. Other genes that were up-regulated in the microarray analyses were several additional hypothetical proteins as well as SGO_1109 involved in pyrimidine synthesis and SGO_1695 involved in fatty acid biosynthesis pathways (Table 1).

The genes that showed the largest negative fold changes in response to hyphae encoded proteins that generally fell into three predicted functional categories (Tanenbaum *et al.*, 2010): energy metabolism (SGO_0184, SGO_0440, SGO_0631, SGO_1283, SGO_1300, SGO_1592, SGO_1593, SGO_0384), central intermediary metabolism (SGO_0278, SGO_1720, SGO_1757), and transport and binding proteins (SGO_0383, SGO_0630, SGO_0982, SGO_0985, SGO_1216). The shifts in expression generally relate to genes involved in amino acid metabolism and transport. Accordingly, representative genes *arcA* and *arcB*, involved in arginine catabolism, and *glmS*, involved in the formation of glutamate from glutamine were chosen for validation of decreased expression in DL1 in response to hyphae by real time PCR (Table 2).

The role of *fruRBA* in strain DL1 biofilm formation

The doubling times of planktonic cultures of the parent strain DL1 and a $\Delta fruRBA$ mutant strain were similar in TY or YPT media independent of either glucose or fructose as a carbohydrate source. In biofilm experiments, cultures were grown to the same OD₆₀₀ as starting inocula, in the respective medium to avoid any nutritional shift effects. The biofilms formed over a 16 h period by the $\Delta fruRBA$ mutant were less dense than those of the parental strain DL1 in TY-G medium (Fig. 1). Although there were no statistically significant differences in biofilm mass based upon crystal violet staining assay, there were distinct differences in the architectural patterns of cells present within the biofilms. The mutant cells tended to form aggregates and these aggregates were even more visually evident in the TY-F medium biofilms (Fig. 1). In YPT-G medium, the $\Delta fruRBA$ mutant formed denser aggregates, whereas parental cells were more evenly dispersed across the substratum. Aggregates of the $\Delta fruRBA$ mutant were also visible in YPT-F medium biofilms (Fig. 1).

Role of *fruRBA* in dual species biofilms

Because the aggregation differences between parent and mutant cells were most visually evident in YPT-G medium, this medium was chosen for additional dual species biofilm studies. Visual examination of light microscopy images (Fig. 2) indicated that biofilms formed by *C. albicans* were enhanced in the presence of *S. gordonii* DL1, covering more of the substratum surface than those biofilms formed by *C. albicans* alone. In contrast, the dual species biofilm formed by *C. albicans* with strain UB2683, the $\Delta fruRBA$ mutant, covered less of the substratum surface and had fewer intertwining hyphal filaments than dual species biofilms formed with the parental strain DL1. Complementing the $\Delta fruRBA$ mutation with plasmid-borne *fruRBA* genes in strain UB2683/p49M*fruRBA* resulted in a reversion of the dual-species biofilm architecture to one more similar to that seen with strain DL1. CLSM images of dual-species biofilms suggested that overall there were fewer $\Delta fruRBA$ mutant cells associated with *C. albicans* hyphal filaments compared with *S. gordonii* DL1-*C. albicans* biofilms (Fig. 3). In the $\Delta fruRBA$ mutant-*C. albicans*

biofilms, many more hyphal filaments were visibly exposed (Fig. 3b) and they extended further into the environment. Consequently the $\Delta fruRBA$ mutant dual-species biofilms were of greater maximum height (see legend to Fig. 3) than the other biofilms in which the hyphal filaments tended to be more closely retained within the mixed species communities (Fig. 3). The complemented mutant produced a dual-species biofilm similar to wild type (Fig. 3c). Thus, the *fruRBA* operon is necessary for normal biofilm formation of *S. gordonii* with *C. albicans*, and is one of the operons up-regulated early in *S. gordonii* under co-culture conditions.

DISCUSSION

Oral streptococci and *C. albicans* can co-habit the same niche on human oral cavity surfaces. Dual species interaction studies have identified phenotypic responses and molecular mechanisms that mediate their interactions in oral biofilms. Communication and cellular interactions among members of multispecies oral biofilms can influence the relative abundance of specific microorganism, and consequently influence the relative health or pathogenic potential of the oral microbiome. Such microbial interactions may be mediated by physical-chemical properties, interactions between cell-associated surface molecules, or via extracellular products such as peptides or diffusible fatty acids (Xu *et al.*, 2014a).

To better understand the intercellular communication from the streptococcal perspective, the initial response of *S. gordonii* cells exposed to *C. albicans* forming hyphal filaments was examined by transcriptome analysis. A 30 minute co-incubation period was chosen since *S. gordonii* gene expression is known to respond to environmental signals during this timeframe (Vickerman *et al.*, 2007). Hyphal cells were employed in these interaction studies since this morphological form predominates in *C. albicans* biofilms. During this co-incubation, there was no evident up-regulation of genes encoding the streptococcal cell -wall anchored surface proteins that have previously been shown to mediate *S. gordonii*-*C. albicans* binding. For

example, *S. gordonii* CshA protein domains (Holmes *et al.*, 1996) and SspB protein (Silverman *et al.*, 2011) have been shown to interact with the *C. albicans* cell surface. However, expression of neither *cshA* nor *sspB* was up-regulated during the 30 min co-incubation indicating that adhesion mediated by these proteins probably does not result from early specific transcriptional responses to *C. albicans* hyphal cells. Although the products of these genes play important role in inter-microbial interactions, their expression may not be specifically induced by the presence of hyphae or their expression may be affected at a later time point in dual-species growth.

Genome-wide transcriptional studies of *S. mutans* after 4 to 24 hour of biofilm growth with *C. albicans* demonstrated up-regulation of genes involved in the alternative competence pathway (Sztajer *et al.*, 2014). This effect was only noted after eight hours of dual species biofilm growth, by which time multiple environmental signals and responses would have occurred affecting multiple gene pathways. Likewise, a role for *S. gordonii* competence genes has been demonstrated in determining the architecture of dual-species biofilms but again, this was seen phenotypically after several hours of co-cultivation (Jack *et al.*, 2015). The competence pathway of *S. gordonii* differs from that of *S. mutans* and is more similar to that of *Streptococcus pneumoniae* with a response that is essentially completed 45 min after induction (Vickerman *et al.*, 2007). In the present studies there were no differentially-expressed competence peptide-responsive genes seen in *S. gordonii* cells co-incubated with *C. albicans* in the 30 min time frame. Taken collectively, the various studies suggest that competence-related responses play a role after the initiation of the dual species biofilms.

In general, the *S. gordonii* genes that initially responded to the presence of hyphae, when compared to pH-matched medium controls, were those involved in carbohydrate and amino acid metabolism. The majority of former were up-regulated whereas the latter were down-regulated. *In silico* analyses using various online sequence comparison programs such as Fuzznuc and Clustal (Rice *et al.*, 2000) did not detect DNA sequences conserved among the

upstream promoter regions of all the up- or down-regulated genes that would be suggestive of potential signal factor binding sites (data not shown). Fold changes in gene expression were modest, but genes were detected with differential expression that was reproducible and statistically significant. Several loci for hypothetical proteins were responsive to hyphae, but their functions are unknown and the potential roles of those genes cannot yet be determined. The most significantly up-regulated hyphal-responsive *S. gordonii* genes were those in the *fruRBA* operon. The *fruR* gene encodes a phosphotransferase system (PTS) repressor, *fruB* encodes a 1-phosphofructokinase, and *fruA* encodes the fructose specific permease of the phosphoenolpyruvate-dependent sugar phosphotransferase system. The phosphotransferase system enzymes in streptococci comprise the main mechanisms for transporting carbohydrates across the membrane into the cell for energy (Pessione, 2012). These carbohydrate transporters show some redundancy in their sugar specificity (Deutscher *et al.*, 2014; Ajdic & Pham, 2007). The genes in the *S. gordonii* fructose PTS operon were originally identified and characterized (Loo *et al.*, 2003) based upon their sequence and functional similarities to the *S. mutans fruRKI* genes. However, since *S. gordonii* is more closely related phylogenetically to *S. pneumoniae*, which was used as the basis for annotation of *S. gordonii* genome sequence (Vickerman *et al.*, 2007), and the *S. pneumoniae* homologs are designated *fruRBA*, in the present paper we refer to these *S. gordonii* genes with the *fruRBA* nomenclature used in the NCBI genome database.

Co-incubation of DL1 cells with heat-killed vs. live hyphae in fresh pH-matched medium confirmed that as-yet-unidentified heat-stable component(s) associated with the living hyphae themselves (e.g. carbohydrates, small peptides) contribute to the *fruRBA* operon response. Reporter gene studies have shown that *fruB* is induced by fructose, sucrose, xylitol and human serum, and that expression of this operon is subject to glucose catabolite repression (Loo *et al.*, 2003). The surfaces of *C. albicans* cells are covered with carbohydrate-containing compounds

such as chitin, mannans and β -glucans (Dongari-Bagtzoglou *et al.*, 2009). These can be hydrolyzed into carbohydrate moieties via various streptococcal and fungal enzymes e.g. N-acetylglucosaminidase, glucosidase, glucanase; and so upregulation of the *fruRBA* operon in response to hyphae is consistent with bacterial sensing alternative carbohydrate availability.

Another intriguing gene that was up-regulated in response to hyphae is the gene encoding the enoyl-acyl carrier protein. Additional fatty acid metabolism genes were up-regulated in response to hyphae but they did not meet our stringent statistical cutoff to be included in Table 1 (data are in the GEO files). Recent studies suggest that diffusible fatty acid signals made by several oral streptococcal species (Vilchez *et al.*, 2010) may be involved in influencing or inhibiting hyphae formation, but it is not known at present if *S. gordonii* produces these. Future studies may provide more insights into the signals and cognate responses of these two microorganisms at various stages of *S. gordonii*-*C. albicans* biofilm initiation and growth.

Small peptides are a common mechanism of streptococcal intercellular signaling and quorum sensing (Kleerebezem *et al.*, 1997). Although there was no down-regulation among genes expressing known *S. gordonii* quorum sensing peptides, a putative bacteriocin accessory protein, SGO_1216, was down-regulated 1.8-fold, consistent with the expression of its homolog in *S. mutans* which was ca. 2- to 5- fold down-regulated in 6-, 10- and 24-h dual species biofilm growth with *C. albicans* (Sztajer *et al.*, 2014). The functional specificity of this transporter remains to be determined, as does its role in *Streptococcus*-*C. albicans* interactions. Nevertheless, this result is intriguing because of the shared down-regulation response of these homologs in two different oral streptococcal species in response to *C. albicans*.

The majority of the *S. gordonii* genes that were down-regulated in an initial response to the environmental presence of *C. albicans* hyphae appeared to indicate metabolic changes. Two main sources of energy in *S. gordonii* and other lactic acid bacteria are sugar

phosphorylation and arginine deimination (Pessione, 2012). Indeed, many of the down-regulated genes encode putative functions in the arginine metabolic pathways (Kloosterman & Kuipers, 2011). Glutamine is centrally involved in both arginine and pyrimidine metabolism pathways. It can be converted to glutamate and involved in the arginine biosynthesis pathways via ornithine and citrulline, or it can be converted to carbamoyl-phosphate with roles in energy production and pyrimidine synthesis (Kloosterman & Kuipers, 2011). The gene encoding GlnS involved in the formation of glutamate from glutamine also has a role in arginine and pyrimidine synthetic pathways (Kloosterman & Kuipers, 2011). Similarly, three genes of the arginine deiminase pathway were down-regulated in response to hyphae. This pathway involves arginine conversion by ArcA to citrulline and ammonia. Citrulline is converted to ornithine or carbamoyl phosphate by Arc B. Carbamoyl phosphate is involved in pyrimidine metabolism, or via ArcC can yield ATP, as well as ammonia to neutralize potential acidic pH. Changes in expression of metabolic genes in response to the initial presentation of the numerous sugars available from *C. albicans* would suggest a switch to carbohydrate utilization, consistent with metabolic pathway-utilization gene or protein changes seen in other Gram-positive bacteria co-cultivated with *C. albicans* (Peters *et al.*, 2010; Sztajer *et al.*, 2014). Overall, our transcriptome analyses were not exhaustive and genes that were differentially expressed below our stringent threshold cutoff levels may also be biologically important. It would be expected that gene expression patterns would change as dual species growth progresses. Nevertheless, the present studies have identified genes that were consistently and reproducibly affected by initial exposure of *S. gordonii* to hyphae, thereby preparing groundwork for future genetic and functional investigations.

Because genes in the *fruRBA* operon showed the most significant up-regulation in response to the presence of hyphae, we further investigated the potential role of these genes in dual-species biofilm formation. Previous studies have shown that *S. gordonii* strains with

mutations in genes in the *fruRBA* operon formed less robust biofilms on abiotic glass, saliva-coated (Loo *et al.*, 2003;) or fibronectin-coated (Bizzini *et al.*, 2006) substrata compared to those of parental cells. Nonpolar mutations in *fruB* and *fruA* resulted in significant reductions of biofilm formation on glass or experimental salivary pellicle, whereas nonpolar disruption of *fruR* had no effect (Loo *et al.*, 2003). The mechanism underlying the reduction in streptococcal biofilm formation is unknown and was speculated to involve additional roles of FruB in two-component signal transduction by acting as a kinase for a different regulatory system via cross-regulation (Deutscher *et al.*, 2014). This possibility is consistent with studies suggesting that PTS's may have global regulatory roles in response to environmental cues (Lengeler & Jahreis, 2009; Pereira *et al.*, 2012). In the present studies, deletion of all three genes of the *fruRBA* operon resulted in changes to the biofilm architecture and surface coverage patterns on saliva-coated glass cover slips. The streptococcal *fruRBA* mutant cells formed more macroscopic aggregates thereby leaving more of the substratum uncovered. Such aggregation of cells may be a community mechanism to keep cells in closer proximity in order to optimize signaling and shared extracellular products that may favor nutrient processing and availability (Rickard *et al.*, 2003).

The experimental design of the dual-species biofilm experiments only allowed streptococcal cells that attached in 30 min to a hyphae-coated substratum to contribute to the mature biofilm formation. Based upon the subsequent growth pattern, the mutant cells tended to attach in clusters suggesting that during the initial attachment phase the cells remained nearer each other rather than being more randomly distributed across the hyphal filament substratum. The similar chain lengths of planktonic parent and mutant cells in the various growth media (data not shown) suggested that this pattern was due to independent cell attachment rather than cell clusters due to cell chaining differences. The difference in parent and mutant biofilms formed during the 30 min attachment period, a time during which the microarray studies

indicated that parental cells up-regulated expression of the *fruRBA* operon, support the idea that these genes play a role in the cellular adhesion patterns during initial interactions with the *C. albicans* cells. Alternatively, it is possible that the phenotypic variation in biofilms of the *fruRBA* mutant strain were due to compensatory expression of other genes in the mutant strain. An examination of the transcriptome of the *S. gordonii fruRBA* mutant either alone or in the presence of hyphae has not been performed. Nevertheless, complementing the mutant strain with plasmid-borne *fruRBA* resulted in a reversion of the biofilm phenotype, confirming a role of the *fruRBA* genes in dual species biofilm formation.

Overall, these studies provide the first genome-wide insights into the initial responses of *S. gordonii* to *C. albicans* hyphae under the conditions described here. The major streptococcal transcriptional effects involve responses to the changed microenvironment and carbohydrate availability. They provide clues to the metabolic responses that result from cross talk between these two microorganisms, and that may go on to influence dual species biofilm development.

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REFERENCES

Ajdić, D. and Pham, V.T. (2007) Global transcriptional analysis of *Streptococcus mutans* sugar transporters using microarrays. *J Bacteriol* **189**: 5049-5059.

Alifano, P., Bruni, C.B., and Carlomagno, M.S. (1994) Control of mRNA processing and decay in prokaryotes. *Genetica* **94**:157-172.

Altschul, S.F., Gish, W., Miller, W., Myers, E.W. and Lipman, D.J. (1990) Basic local alignment search tool. *J Mol Biol* **215**: 403-410.

Bamford, C.V., d'Mello, A., Nobbs, A.H., Dutton, L.C., Vickerman, M.M. and Jenkinson, H.F. (2009) *Streptococcus gordonii* modulates *Candida albicans* biofilm formation through intergeneric communication. *Infect Immun* **77**: 3696–3704.

Bertolini, M.M., Xu, H., Sobue, T., Nobile, C.J., Del Bel Cury, A.A. and Dongari-Bagtzoglou, A. (2015) Candida-streptococcal mucosal biofilms display distinct structural and virulence characteristics depending on growth conditions and hyphal morphotypes. *Mol Oral Microbiol* Mar 7. doi:10.1111/omi.12095 [Epub ahead of print].

Bizzini, A., Beggah-Moller, S., Moreillon, P. and Entenza, J.M. (2006) Lack of *in vitro* biofilm formation does not attenuate the virulence of *Streptococcus gordonii* in experimental endocarditis. *FEMS Immuno Med Microbiol* **48**: 419–423.

Breitling, R., Armengaud, P., Amtmann, A. and Herzyk, P. (2004) Rank products: a simple, yet powerful, new method to detect differentially regulated genes in replicated microarray experiments. *FEBS Lett* **573**: 83–92.

Deutscher, J., Aké, F.M., Derkaoui, M. *et al.* (2014) The bacterial phosphoenolpyruvate: carbohydrate phosphotransferase system: regulation by protein phosphorylation and phosphorylation-dependent protein-protein interactions. *Microbiol Mol Biol Rev* **78**: 231-256.

Diaz, P.I., Xie, Z., Sobue, T. *et al.* (2012) Synergistic interaction between *Candida albicans* and commensal oral streptococci in a novel in vitro mucosal model. *Infect Immun* **80**: 620-632.

Dongari-Bagtzoglou, A., Kashleva, H., Dwivedi, P., Diaz, P. and Vasilakos, J. (2009) Characterization of mucosal *Candida albicans* biofilms. *PLoS ONE* **4**: e7967.

Dutton, L.C., Nobbs, A.H., Jepson, K. *et al.* (2014) O-mannosylation in *Candida albicans* enables development of interkingdom biofilm communities. *MBio* **5**: e00911.

Falsetta, M.L., Klein, M.I., Colonne, P.M. *et al.* (2014) Symbiotic relationship between *Streptococcus mutans* and *Candida albicans* synergizes virulence of plaque biofilms in vivo. *Infect Immun* **82**: 1968-1981.

Holmes, A.R., McNab, R. and Jenkinson, H.F. (1996) *Candida albicans* binding to the oral bacterium *Streptococcus gordonii* involves multiple adhesin-receptor interactions. *Infect Immun* **64**: 4680-4685.

Holmes, A.R. and Shepherd, M.G. (1988) Nutritional factors determine germ tube formation in *Candida albicans*. *J Med Vet Mycol* **26**: 127-131.

Jack, A.A., Daniels, D.E., Jepson, M.A. *et al.* (2015) The *Streptococcus gordonii* *comCDE* (competence) operon modulates biofilm formation with *Candida albicans*. *Microbiology* **161**: 411-421.

Jakubovics, N.S., Strömberg, N., van Dolleweerd, C.J., Kelly, C.G. and Jenkinson, H.F. (2005) Differential binding specificities of oral streptococcal antigen I/II family adhesins for human or bacterial ligands. *Mol Microbiol* **55**: 1591-1605.

Jenkinson, H.F. (2011) Beyond the oral microbiome. *Environ Microbiol* **13**: 3077-3087.

Jenkinson, H.F., Lala, H.C. and Shepherd, M.G. (1990) Coaggregation of *Streptococcus sanguis* and other streptococci with *Candida albicans*. *Infect Immun* **58**: 1429-1436.

Kleerebezem, M., Quadri, L.E.N., Kuipers O.P. and de Vos W.M. (1997). Quorum sensing by peptide pheromones and two-component signal-transduction systems in Gram-positive bacteria. *Mol Microbiol* **24**: 895-904.

Kloosterman T.G. and Kuipers O.P. (2011). Regulation of arginine acquisition and virulence gene expression in the human pathogen *Streptococcus pneumoniae* by transcription regulators ArgR1 and AhrC. *J Biol Chem*. **286**: 44594-605.

Kolenbrander, P.E., Palmer, R.J. Jr., Periasamy, S. and Jakubovics, N.S. (2010) Oral multispecies biofilm development and the key role of cell-cell distance. *Nat Rev Microbiol* **8**: 471-480.

Koressaar, T. and Remm, M. (2007) Enhancements and modifications of primer design program Primer3. *Bioinformatics* **23**: 1289-1291.

Lengeler, J.W. and Jahreis, K. (2009) Bacterial PEP-dependent carbohydrate:phosphotransferase systems couple sensing and global control mechanisms. *Contrib Microbiol* **16**: 65-87.

Livak, K.J. and Schmittgen, T.D. (2001) Analysis of relative gene expression data using real-time quantitative PCR and the $2(\Delta-\Delta C(T))$ Method. *Methods* **25**: 402-408.

Loo, C.Y., Mittrakul, K., Voss, I.B., Hughes, C.V. and Ganeshkumar, N. (2003) Involvement of an inducible fructose phosphotransferase operon in *Streptococcus gordonii* biofilm formation. *J Bacteriol* **185**: 6241–6254.

Macrina, F.L., Tobian, J.A., Jones, K.R. and Evans, R.P. (1981) Molecular cloning in the streptococci In *Genetic Engineering of Microorganisms for Chemicals*. Hollaender, A., DeMoss, R., Kaplan, S., Konisky, J., Savage, D. and Wolfe, R. (eds). New York: Plenum, pp. 195-210.

Morales, D.H. and Hogan, D.A. (2010) *Candida albicans* interactions with bacteria in the context of human health and disease. *PLoS Pathog* **6**: e1000886.

Nobbs, A.H. and Jenkinson, H.F. (2015) Interkingdom networking within the oral microbiome. *Microbes Infect* Mar 21 doi:10.1016/j.micinf.2015.03.008 [Epub ahead of print].

Pereira, C.S., Santos, A.J.M., Bejerano-Sagie, M., Correia, P.B., Marques, J.C. and Xavier, K.B. (2012) Phosphoenolpyruvate phosphotransferase system regulates detection and processing of the quorum sensing signal autoinducer-2. *Mol Microbiol* **84**: 93-104.

Pessione, E. (2012) Lactic acid bacteria contribution to gut microbiota complexity: lights and shadows. *Front Cell Infect Microbiol* **2**: 86. doi: 10.3389/fcimb.2012.00086.

Peters, B.M., Jabra-Rizk, M.A., Scheper, M.A., Leid, J.G., Costerton, J.W. and Shirtliff, M.E. (2010) Microbial interactions and differential protein expression in *Staphylococcus aureus*-*Candida albicans* dual-species biofilms. *FEMS Immunol Med Microbiol* **59**: 493–503.

Podbielski, A., Spellerberg, B., Woischnik, M., Pohl, B. and Lutticken, R. (1996) Novel series of plasmid vectors for gene inactivation and expression analysis in group A streptococci (GAS). *Gene* **177**: 137-147.

Quackenbush, J. (2002) Microarray data normalization and transformation. *Nature Genetics Supplement* **32**: 496-501.

Rice, P., Longden, I. and Bleasby, A. (2000) EMBOSS: The European Molecular Biology Open Software Suite. *Trends Genet* **16**: 276-277.

Rickard, A.H., Gilbert, P., High, N.J., Kolenbrander, P.E. and Handley, P.S. (2003) Bacterial coaggregation: an integral process in the development of multi-species biofilms. *Trends Microbiol* **11**: 94-100.

Saeed, A.I., Sharov, V., White, J. *et al.* (2003) TM4: A Free, Open-Source System for Microarray Data Management and Analysis. *BioTechniques* **34**: 374-378.

Silverman, R.J., Nobbs, A.H., Vickerman, M.M., Barbour, M.E. and Jenkinson, H.F. (2010) Interaction of *Candida albicans* cell wall Als3 protein with *Streptococcus gordonii* SspB adhesin promotes development of mixed-species communities. *Infect Immun* **78**: 4644-4652.

Sztajer, H., Szafranski, S.P., Tomasch, J. *et al.* (2014) Cross-feeding and interkingdom communication in dual-species biofilms of *Streptococcus mutans* and *Candida albicans*. *ISME J* **8**: 2256–2271.

Tanenbaum, D.M., Goll, J., Murphy, S. *et al.* (2010) The JCVI standard operating procedure for annotating prokaryotic metagenomic shotgun sequencing data. *Stand Genomic Sci* **2**: 229–237.

Tatusova, T., Ciufo, S., Fedorov, B., O'Neill, K. and Tolstoy, I. (2014) RefSeq microbial genomes database: new representation and annotation strategy. *Nucleic Acids Res* **42**(Database issue): D553-9.

Untergrasser, A., Cutcutache, I., Koressaar, T. *et al.* (2012) Primer3 - new capabilities and interfaces. *Nucleic Acids Res* **40**: e115.

Vickerman, M.M., Wang, M. and Baker, L.J. (2003) An amino acid change near the carboxyl terminus of the *Streptococcus gordonii* regulatory protein Rgg affects its abilities to bind DNA and influence expression of the glucosyltransferase gene *gtfG*. *Microbiology* **149**: 399-406.

Vickerman, M.M., Iobst, S., Jesionowski, A.M., and Gill, S.R. (2007) Genome-wide transcriptional changes in *Streptococcus gordonii* in response to competence signaling peptide. *J Bacteriol* **189**: 7799-7807.

Vilchez, R., Lemme, A., Ballhausen, B. *et al.* (2010) *Streptococcus mutans* inhibits *Candida albicans* hyphal formation by the fatty acid signaling molecule trans-2-decenoic acid (SDSF). *Chembiochem* **11**: 1552-1562.

Wright, C.J., Burns, L.H., Jack, A.A. *et al.* (2013) Microbial interactions in the building of communities. *Mol Oral Microbiol* **28**: 83-101.

Xu, H., Jenkinson, H.F. and Dongari-Bagtzoglou, A. (2014a) Innocent until proven guilty: mechanisms and roles of *Streptococcus-Candida* interactions in health and disease. *Mol Oral Microbiol* **29**: 99-116.

Xu, H., Sobue, T., Thompson, A. *et al.* (2014b) Streptococcal co-infection augments *Candida* pathogenicity by amplifying the mucosal inflammatory response. *Cell Microbiol* **16**: 214-231.

Zuker M. (2003) Mfold web server for nucleic acid folding and hybridization prediction. *Nucleic Acids Res.* **31**: 3406-3415.

Supporting Information.

Additional Supporting Information may be found in the online version of this article:

Table S1. Oligonucleotide primers used.

Table S2. Comparison of *S. gordonii* CH1 genome loci annotation for CP000725.1 and

NC_009785.

Figure S1. *S. gordonii fruRBA* chromosomal region and DNA used for strain construction.

Table 1. *S. gordonii* genes differentially expressed by microarray analysis

| Locus ^a | Description ^b | Symbol | Predicted Functional Category/Role ^c | Fold Change ^d | P Value ^e |
|---------------------|---|--------|--|--------------------------|----------------------|
| Up-regulated | | | | | |
| SGO_1112 | 1-phosphofructokinase | fruB | Energy metabolism/Glycolysis_gluconeogenesis | 4.0 | 0.002 |
| SGO_1111 | Phosphotransferase system repressor | fruR | Energy metabolism/Biosynthesis and degradation of polysaccharides | 3.4 | 0.002 |
| SGO_1113 | PTS system, fructose specific IIABC components | fruA | Regulatory functions/Other | 3.4 | 0.001 |
| SGO_0702 | hypothetical protein | | Hypothetical | 3.1 | 0.002 |
| SGO_1109 | aspartate carbamoyltransferase catalytic subunit | pyrB | Purines, pyrimidine, nucleosides, and nucleotides/Pyrimidine ribonucleotide biosynthesis | 2.0 | 0.048 |
| SGO_1190 | hypothetical protein | | Hypothetical | 1.6 | 0.029 |
| SGO_1244 | hypothetical protein | | Hypothetical | 1.6 | 0.026 |
| SGO_1686 | hypothetical protein | | Hypothetical | 1.6 | 0.021 |
| SGO_1695 | enoyl-acyl carrier protein (ACP) reductase | | Fatty acid and phospholipid metabolism/Biosynthesis | 1.6 | 0.027 |
| SGO_1696 | hypothetical protein | | Hypothetical | 1.6 | 0.014 |

Down-regulated^f

| | | | | | |
|----------|---|------|---|------|-------|
| SGO_1592 | ornithine carbamoyltransferase | arcB | Energy metabolism/Amino acids and amines | -2.2 | 0.034 |
| SGO_0383 | proton/sodium- glutamate symport protein | | Transport and binding proteins/Amino acids, peptides and amines | -2.0 | 0.002 |
| SGO_1301 | hypothetical protein | | Hypothetical | -2.0 | 0.003 |
| SGO_0440 | L-itol 2-dehydrogenase | | Energy metabolism/Sugars | -2.0 | 0.019 |
| SGO_1593 | arginine deiminase | arcA | Energy metabolism/Amino acids and amines | -1.9 | 0.009 |
| SGO_1757 | glucosamine-fructose-6-phosphate aminotransferase | glmS | Central intermediary metabolism/Amino sugars | -1.9 | 0.008 |
| SGO_1300 | NADPH-dependent reductase | | Energy metabolism/Electron transport | -1.8 | 0.005 |
| SGO_0985 | amino acid permease protein SP0711 | | Transport and binding proteins/Amino acids, peptides and amines | -1.8 | 0.01 |
| SGO_1216 | putative bacteriocin transport accessory protein | | Transport and binding proteins/Unknown substrate | -1.8 | 0.007 |
| SGO_0184 | urease cluster protein | | Energy metabolism/Electron transport | -1.8 | 0.008 |
| SGO_0384 | putative carboxylate-amine/thiol ligase | | Energy metabolism/Other | -1.8 | 0.008 |
| SGO_0631 | alpha-glycerophosphate oxidase | | Energy metabolism/Other | -1.8 | 0.007 |
| SGO_1720 | Aminotransferase, class-V | | Central intermediary metabolism_Amino acid | -1.8 | 0.011 |

| | | | | | |
|----------|--|------|---|------|-------|
| | | | biosynthesis/Other_Aspartate family | | |
| SGO_0630 | glycerol uptake facilitator protein | | Transport and binding proteins/Other | -1.7 | 0.012 |
| SGO_0278 | peptide methionine sulfoxide reductase | msrA | Central intermediary metabolism/Other | -1.7 | 0.01 |
| SGO_0982 | amino acid transport protein | | Transport and binding proteins/Amino acids, peptides and amines | -1.6 | 0.03 |
| SGO_1283 | pyridine nucleotide-disulfide oxidoreductase | | Energy metabolism/Electron transport | -1.6 | 0.043 |

^a Locus tag in *S. gordonii* genome sequence for CP000725.1 (Vickerman *et al.*, 2007).

^b Locus description in NCBI database annotation for CP000725.1.

^c Predicted Functional Category/Role (Tanenbaum *et al.*, 2010)

^d Average fold change, based upon cDNA probes hybridizing to a minimum of 24 up to a maximum of 48 independent 70-oligomer spots for each locus, of DL1 cells co-incubated with hyphae compared to baseline values of DL1-CSB-Glc pH-matched controls in two biologically independent experiments.

^e P value is based upon the rank product algorithm (Breitling *et al.*, 2004) of the MEV software.

^f A negative sign added to the absolute fold change value for the downregulated genes indicates the value for DL1 cells incubated with hyphae is less than the DL1-CSB-Glc value.

Table 2. Quantitative PCR validation of gene expression during *S. gordonii* and hyphal *C. albicans* interactions.

| Locus | Gene Name | Relative Expression (2 ^{-(ΔΔC_t)}) ^a |
|---|--------------|---|
| | | Avg. Expression |
| Potentially Up-regulated by Microarray analysis | | |
| SGO_1111 | <i>fruR</i> | 6.5 ± 1.98 |
| SGO_1112 | <i>fruB</i> | 9.05 ± 0.64 |
| SGO_1113 | <i>fruA</i> | 3.75 ± 0.92 |
| SGO_0701 | <i>hup</i> | 0.95 ± 0.21 |
| SGO_0702 | Hypothetical | 1.95 ± 0.35 |
| Potentially Down-regulated by Microarray analysis | | |
| SGO_1593 | <i>arcA</i> | 0.6 ± 0.28 |
| SGO_1592 | <i>arcB</i> | 0.6 ± 0.28 |
| SGO_1757 | <i>glmS</i> | 0.5 ± 0.14 |

^a Values were normalized to 150- or 100-bp amplicons of *gyrA* as appropriate. Expression values shown are those of DL1 cells in the hyphal interaction compared to baseline values (set at 1) of DL1 cells in pH-matched CSB-Glc. Relative expression values greater than 1 indicate up-regulation; values less than 1 indicate down-regulation. N=6.

Table 3. Specificity of the *S. gordonii fruRBA* response to *C. albicans* hyphae

| Co-incubations in CSB | Average Relative Expression ($2^{-\Delta\Delta Ct}$) ^a |
|-------------------------------|---|
| <i>fruR</i> | |
| DL1 + hyphae | 9.6 ± 4.9 |
| DL1 + heat-killed hyphae | 2.8 ± 1.5 |
| DL1 + <i>E. faecalis</i> OG1X | 1.6 ± 0.8 |
| <i>fruB</i> | |
| DL1 + hyphae | 12.1 ± 1.3 |
| DL1 + heat-killed hyphae | 3.7 ± 0.3 ^{*b} |
| DL1 + <i>E. faecalis</i> OG1X | 0.8 ± 0.5 [*] |
| <i>fruA</i> | |
| DL1 + hyphae | 18.9 ± 4.0 |
| DL1 + heat-killed hyphae | 3.1 ± 2.0 [*] |
| DL1 + <i>E. faecalis</i> OG1X | 3.8 ± 2.5 [*] |

^a All values normalized to 150-bp amplicons of *gyrA* and compared to the baseline expression of DL1 cells alone.

^b values with asterisks differ significantly from DL1 + hyphae ($p < 0.05$). Significance patterns are consistent with 5-prime to 3-prime net directionality of prokaryotic transcript decay (Alifano *et al.* 1994). $N \geq 12$.

FIGURE LEGENDS

Figure 1. Biofilms formed on saliva-coated glass cover slips. Sixteen hour biofilms of *S. gordonii* DL1 or $\Delta fruRBA$ grown in TY-G or TY-F media (top panels), and in YPT-G or YPT-F media (bottom panels). Two representative views of transmitted light microscopy fields of crystalviolet - stained biofilms formed in each medium are shown for each strain.

Figure 2. Images of single- or dual-species biofilm formation in YPT-G medium by *S. gordonii* cells on saliva-coated cover slips with or without attached *C. albicans* SC5314 cells. Representative light microscopic fields of crystal violet-stained biofilms are shown. The middle panels show *S. gordonii* biofilms of the parent (top), $\Delta fruRBA$ mutant (middle) and *fruRBA* complemented (bottom) strains (DL1, UB2683, and UB2683/p49M*fruRBA*, respectively) grown from cells that bound during a 30-min incubation period to substrata with attached *C. albicans* cells. The flanking panels show corresponding monospecies biofilms of *S. gordonii* (left panels) grown from cells that bound saliva-coated substrata over 30-min incubation, and *C. albicans* biofilm (right panel) grown from cells that attached during 1-h incubation. Magnification Bar = 50 μm .

Figure 3. CLSM images of dual-species *C. albicans* SC5314-*S. gordonii* biofilms formed on saliva-coated glass cover slips in YPT-G medium. Biofilms were grown as described in Methods for 6 h at 37°C and fluorescently stained with FITC. For each of the panels (A-D) a series of x-y sections were collected to generate 3D reconstructions of representative CLSM images with Volocity® software. Heights were measured across ten CLSM image stacks from two independent experiments \pm SD. Panels: (a), *C. albicans*-*S. gordonii* DL1, height $81.4 \pm 7.50 \mu\text{m}$; (b), *C. albicans*-*S. gordonii* $\Delta fruRBA$, height $113 \pm 10.4 \mu\text{m}^*$; (c), *C. albicans*-*S. gordonii* $\Delta fruRBA/pfruRBA$, height $70.6 \pm 6.40 \mu\text{m}$; (d), *C. albicans*, height 86.8 ± 11.6 . * $P < 0.001$ when

heights were compared using the *t* test. Note that in (b) there is more substratum vacant and more hyphae are visible compared with the other dual-species biofilm panels (a) and (c).

Figure S1. Map of DNA used for construction of $\Delta fruRBA$ mutant and complement strains. The chromosomal region of strain DL1 is shown with large arrows showing the positions and directions of the *fruRBA* genes. Software searches identified putative promoters (P) and transcriptional terminators (lollipops) in this region. The ΔG values for putative transcriptional terminators are shown as calculated with mfold (Zuker, 2003; <http://mfold.rna.albany.edu>). Positions of the Fru.F1/FruR1 and FruF2/FruR2 amplicons are shown above their corresponding genomic locations and include the upstream *S. gordonii* gene locus SGO_1110 and the downstream locus SGO_1114, respectively. Numbers directly below the ends of the amplicons (1153026 and 1158017) correspond to the nucleotide numbers in the *S. gordonii* Challis genome sequence (Genbank CP000725.1). The lower diagram shows the linear DNA fragment used to transform strain DL1 to construct strain UB2683, the $\Delta fruRBA$ deletion mutant in which the upstream and downstream amplicons flank an *aad9* determinant with its own promoter and terminator. The upper panel shows the position of the amplicon made with primers BamFruCompF and BamFruCompR that was cloned into a replicative plasmid to generate p49M*fruRBA* used to complement strain UB2683.

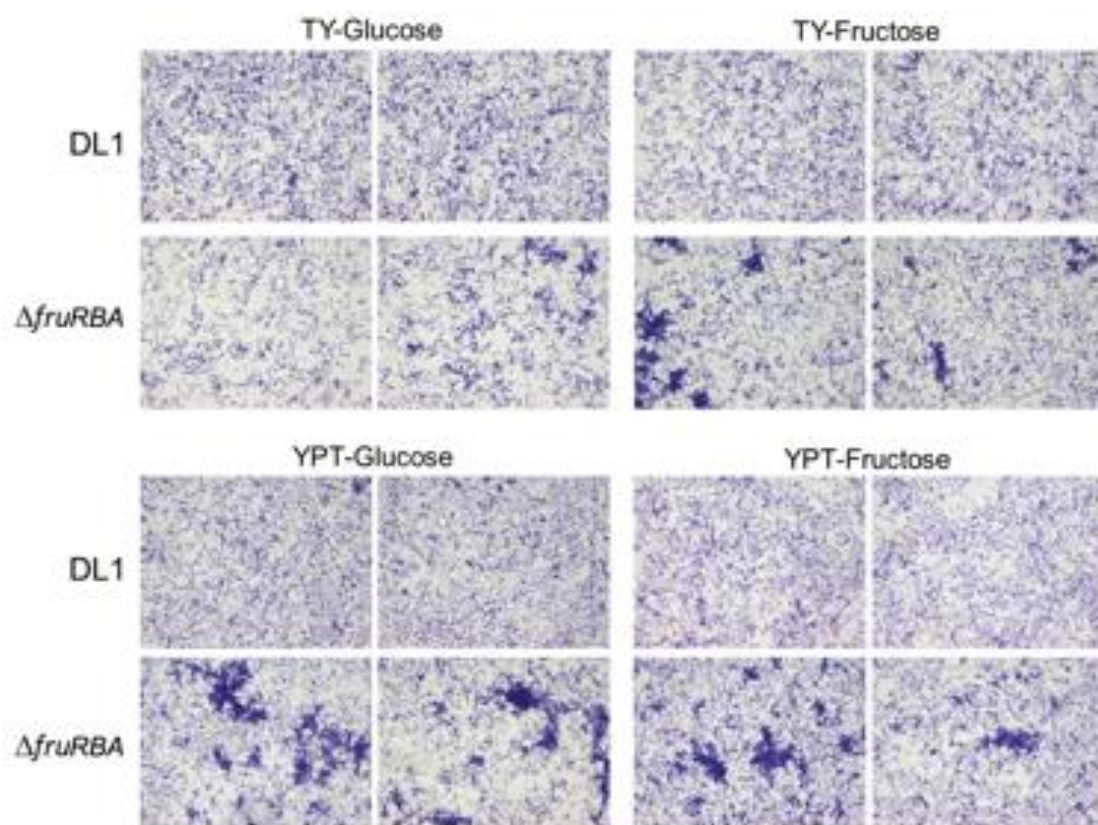


Figure 1

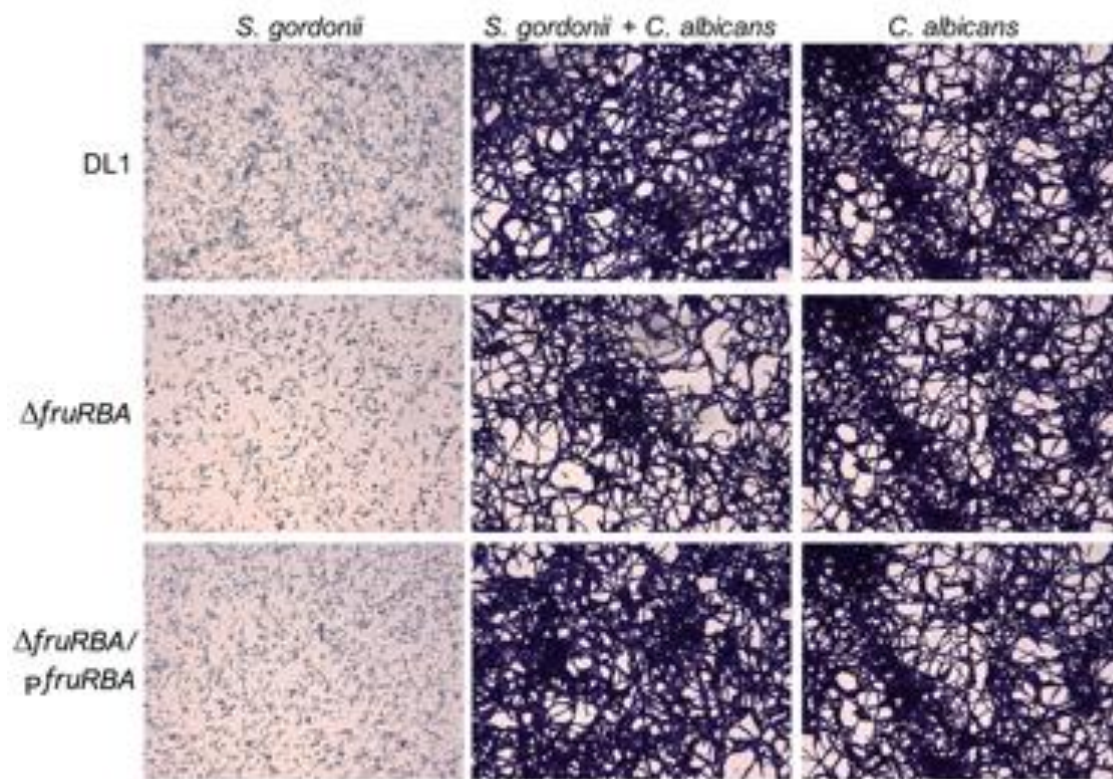


Figure 2

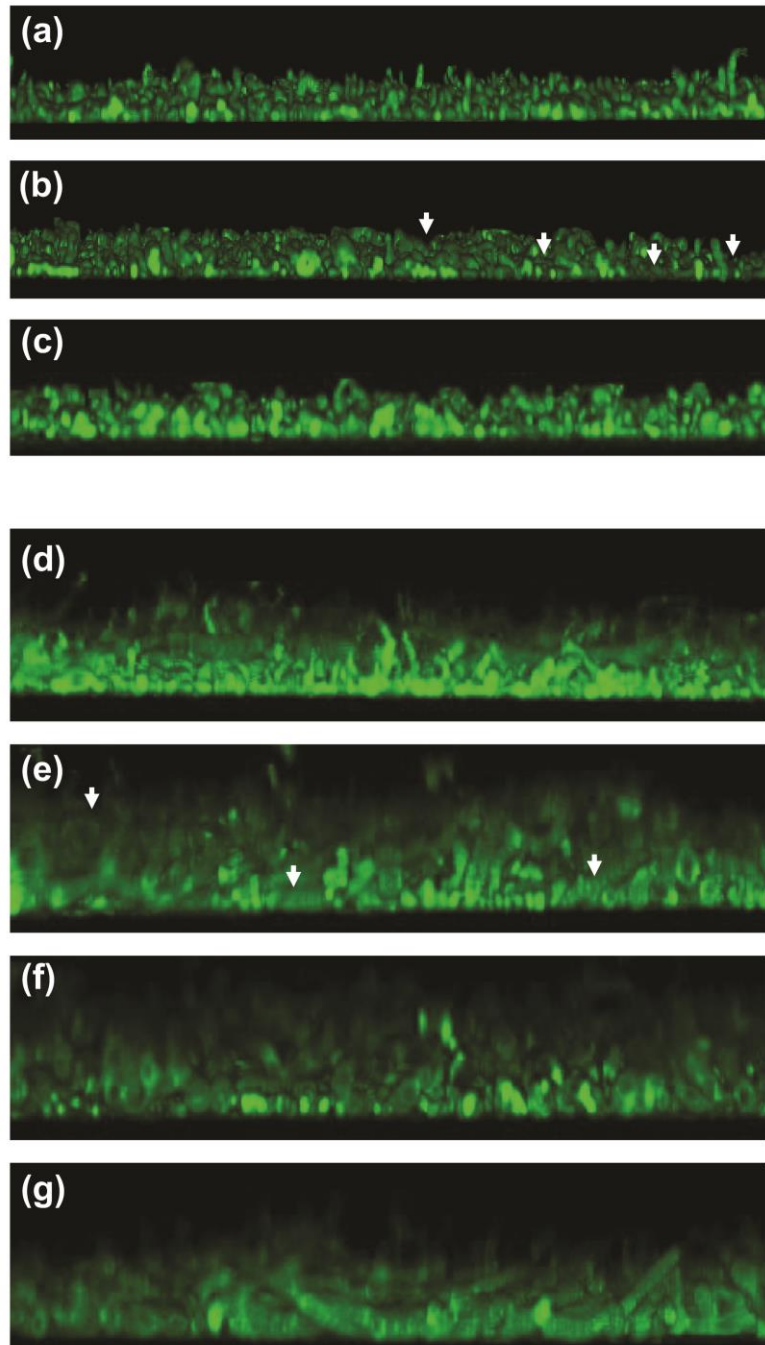


Figure 3

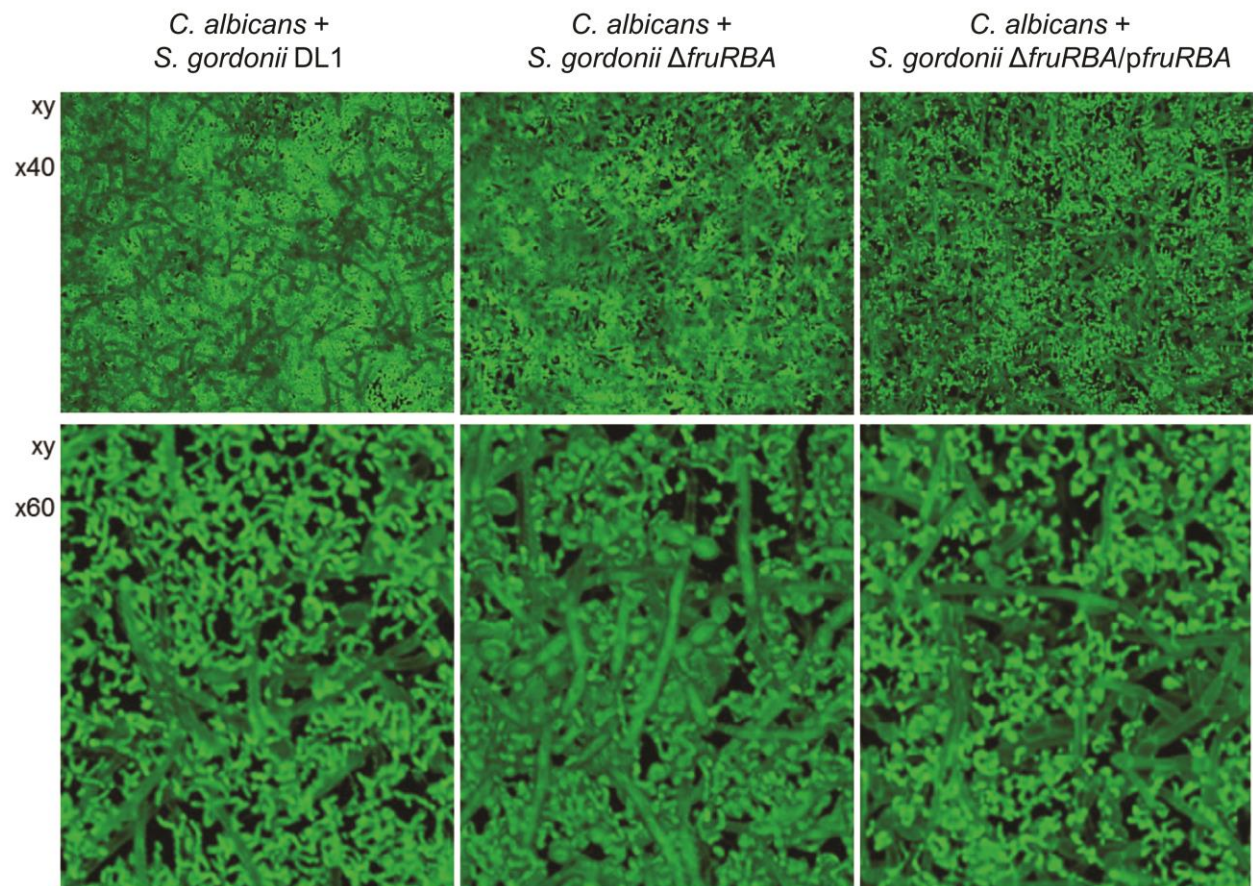


Figure 4